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(71) Applicant: Société des Produits Nestlé S.A.
1800 Vevey (CH)

(72) Inventors:

- Arigoni, Fabrizio
1204 Geneva (CH)
- Schell, Mark Alan
Athens, GA 30605 (US)
- Blum, Stéphanie
1005 Lausanne VD (CH)

- Schiffelin, Eduardo
1023 Crissier VD (CH)
- Delley, Michele
1010 Lausanne (CH)

(74) Representative: Becker Kurlig Straus
Patentanwälte
Bavariastrasse 7
80336 München (DE)

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(54) Serpin from Bifidobacteria

(57) The present invention pertains to a novel gene of Bifidobacteria and to the polypeptides encoded thereby. In particular, the present invention pertains to a gene belonging to the Serpin superfamily and its use in the

production of bacterial Serpins. Also provided are vectors, host cells, and methods for producing bacterial Serpin polynucleotides and/or polypeptides.

Description

[0001] The present invention pertains to a novel gene of Bifidobacteria and to the polypeptides encoded thereby. In particular, the present invention pertains to a gene belonging to the Serpin superfamily and its use in the production of bacterial Serpins. Also provided are vectors, host cells, and methods for producing bacterial Serpin polynucleotides and/or polypeptides.

[0002] Lactic acid bacteria have been utilized for the preservation and preparation of food material for long time taking benefit of the low pH and the action of products generated during the fermentative activity thereof. In addition, Lactic acid bacteria are involved in the production of a variety of different food products, such as cheese or yogurt.

[0003] Quite recently lactic acid bacteria, in particular Lactobacilli and Bifidobacteria, have attracted a great deal of attention in that some strains have been found to exhibit valuable properties to man and animals upon ingestion. These strains, which are generically designated probiotics, have been found to be capable to survive the severe environmental conditions prevailing in the gastric tract and be able to at least transiently colonize the intestinal mucosa, where they bring about positive effects for the living beings having incorporated them.

[0004] In EP 0 768 375 such a probiotic strain of the genus Bifidobacterium is disclosed, which is capable to become implanted in the intestinal flora. This Bifidobacterium is reported to assist in the immuno-modulation of the host, being able to competitively exclude adhesion of pathogenic bacteria to intestinal cells, thus supporting the maintenance of the individual's health.

[0005] Further, in EP 0 577 903 reference is made to the use of a lactic acid bacteria having the ability of replacing *Helicobacter pylori*, the acknowledged cause for the development of ulcer.

[0006] Also, in WO 97/00078 a specific lactobacillus strain, termed *Lactobacillus GG* (ATCC 53103), is disclosed as such a probiotic. The microorganism may be employed for preventing or treating food induced hypersensitivity reactions.

[0007] In view of the valuable properties these probiotic strains may provide, there is a desire for obtaining more detailed information about the biology of these strains, especially about the interaction with the hosts, the phenomena of surviving different environmental conditions in the gut as well as about the capability to adhere to the intestine's mucosa. In particular the involvement thereof in the enhancement of the immune system and defense against pathogens is of high interest.

[0008] Consequently, a problem of the present invention is to provide data about bacterial strains that exhibit properties beneficial for man and/or animals and occasionally elucidate.

[0009] In the line of investigating the genome of the probiotic Bifidobacterium strain BL29 the present inventors have surprisingly found a gene that shows a moderate homology to genes belonging to the Serpin superfamily (SERine Protease Inhibitors). Genes for such type of genes have so far only been found in cells of higher organisms, such as humans and plants, but not in bacterial cells.

[0010] In consequence, the present invention provides for a nucleic acid as identified by SEQ ID. NO. 1 or parts or variants thereof coding for a functional polypeptide, which variants have a homology to the SEQ ID. No. 1 of about 75 %, preferably 80 %, more preferably 85 %, even more preferably 90 % even more preferred 95 %.

[0011] According to an alternative embodiment the present invention also pertains to a polypeptide as identified by SEQ ID. NO. 2 or functional parts or variants thereof, which variants have a degree of homology to the said SEQ. ID. No. 2 of about 75 %, preferably 80 %, more preferably 85 %, even more preferably 90 % even more preferred 95 %.

[0012] Serine proteinase inhibitors (Serpins) comprise a diverse group of proteins that form a superfamily including more than 100 members. The majority of Serpins act as protease inhibitors and are involved in the regulation of several proteinase-activated physiological processes, important for the individual, such as blood clotting, complement mediated lysis, the immune response, glomerulonephritis, pain sensing, inflammation, pancreatitis, cancer, regulating fertilization, bacterial infection and viral maturation. Though the primary function of Serpins appears to be neutralizing serine proteinase activity, these polypeptides have also been found to play a role in extracellular matrix remodelling and cell migration.

[0013] Examples for Serpins include, α 1-antitrypsin, antithrombin III, plasminogen activator inhibitor 1 (PAI-1) or plasminogen activator inhibitor 2.

[0014] The Serpins known so far have been the subject of intensive Research and they all seem to have a common characteristic loop, termed the reactive site loop (RSL), extending from the surface of the molecule containing the recognition sequence for the active site of the cognate serine protease. The specificity of each inhibitor is considered to be determined primarily by the identity of the amino acid that is immediately amino-terminal to the site of potential cleavage of the inhibitor by the serine protease. This amino acid, known as the Pi site residue, is considered to form an acyl bond with the serine in the active site of the serine protease.

[0015] The Serpins seem to act as "suicide inhibitors" forming a 1 : 1 stoichiometric complex with the target proteinase, thus blocking their activity. According to recent data it has been indicated that the inhibitor is cleaved in the reactive center and that the complex is most likely trapped as a covalent acyl-enzyme complex.

[0016] Since Serpins are involved in sophisticated biological processes such as modulating the immune system or inflammatory reactions or even remodelling the extracellular matrix of a higher living being their presence in prokaryotes has not been expected. In fact, no Serpin has so far been reported to be derived from prokaryotic cells.

[0017] The present inventors are therefore the first to have found that also some bacterial cells may contain Serpins. 5 Without wishing to be bound by any theory it is presently assumed that the probiotic properties, such as modulation of the immune system or the known property of the Bifidobacterial strain, from which it has been derived, may at least in part be due to the presence of the present Serpin.

[0018] In the context of this application a nucleic acid according to the present invention shall designate a polynucleotide as identified by SEQ. ID. NO. 1 or parts or variants thereof, that yield a functional polypeptide. To this end, the 10 nucleic acid shown in SEQ ID NO. 1 may be truncated at its ends to an extent, at which the resulting polypeptide still yields the biological function. Likewise, the nucleic acid may also be modified by deleting, adding or replacing one or more nucleotides, with the proviso that the resulting polypeptide still exerts its biological function.

[0019] Similarly, the same applies to the polypeptide described herein. The term polypeptide according to the present 15 invention shall designate a polypeptide as identified by SEQ. ID. NO. 2 or parts or variants thereof, that are functional. Therefore, the polypeptide of SEQ. ID. NO. 2 may be truncated at its ends to an extent, at which the polypeptide still yields the biological function. Likewise, the polypeptide may also be modified by having one or more amino acids being deleted, added or replaced, with the proviso that the resulting polypeptide still exerts its biological function.

[0020] According to an embodiment, the above mentioned nucleic acid may be inserted in a suitable host cell and 20 expressed therein. For this purpose, a nucleic acid according to the present invention may be inserted in a suitable vector, which allows propagation and/or expression in the desired host cell and inserted therein. The vector will contain a marker gene to enable a stable propagation.

[0021] Likewise, the nucleic acid according to the present invention may also be included into the genome of the 25 host, using the phenomenon of homologous recombination or other techniques, allowing insertion of a nucleic acid only into the host's chromosome. Such a technique is e.g. described in EP 93 105 303.7, the contents thereof is incorporated herein by way of reference.

[0022] The nucleic acid according to the present invention may be put under the control of an endogeneous or 30 exogeneous regulon, e.g. a promotor, depending on whether the gene product shall be over-expressed, i.e. for collecting and purifying the polypeptide, or expressed to a certain extent to be delivered to an individual via a carrier system, such as a micro-organism. The regulon, e.g. the promoter, is preferably regulatable and/or inducible and will be operably linked with the coding molecule via any of the well-recognized and easily-practised methodologies for so doing.

[0023] These recombinant constructs are then introduced for expression into suitable host cells such as, e.g., E. 35 coli, Lactobacilli, Streptococci or Bifidobacteria as a prokaryotic host cell or Saccharomyces cerevisiae, insect cells, CHO or COS cells as eukaryotic host cells and the transformed or transduced host cells are cultured under conditions which allow expression of the heterologous gene. It will be appreciated that the gene product of the present polynucleotide will be subject to glycosylation upon expression in an eucaryotic expression system.

[0024] According to another aspect the present invention also comprises recombinant micro-organisms containing 40 at least a copy of the nucleic acid according to the present invention. The nucleic acid may be included in a micro-organism, that is used to deliver the target-substance to the individual. In this respect probiotic bacteria are suitable, since they are able to pass the gastric tract of an individual and get at least transiently implanted into the mucosa of a host. At this location it will exert its biological function as is seen in the present probiotic strain BL29, from which it was derived. Without wishing to be bound by any theory it is presently thought that the gene product of the present nucleic acid is involved in the anti-inflammatory activity displayed by the Bifidobacterial strain BL29. Likewise, any bacterial strain, already including a nucleic acid according to the present invention may be used as a host cell, into 45 which additional copies of the target nucleic acid may be included.

[0025] The host cell will express the gene product of a nucleic acid according to the present invention. Therefore, the host cells may be utilized for the synthesis of a polypeptide according to the present invention on large scale.

[0026] A "host cell" according to the present invention may be obtained by recombinant means, in that a nucleic acid 50 according to the present invention is inserted in a suitable cell. However, in order to increase the amount of Bifido-Serpins in the Bifidobacteria containing such sort of polypeptides, the Bifidobacteria itself may be subjected to common techniques of mutation or selection such that a strain having an increased amount of the corresponding gene product is obtained.

[0027] The isolation of the protein can be carried out according to known methods from the host cell or from the 55 culture supernatant of the host cell. Such methods are described for example by Ausubel I., Frederick M., Current Protocols in Mol. Biol. (1992), John Wiley and Sons, New York. The polypeptide can be purified after recombinant production by affinity chromatography using known protein purification techniques, including immunoprecipitation, gel filtration, ion exchange chromatography, chromatofocussing, isoelectric focussing, selective precipitation, electrophoresis, or the like.

[0028] The invention further comprises a method for detecting a nucleic acid or a polypeptide according to the present invention, comprising incubating a sample, e.g. cell lysates or a reverse transcript of an RNA sample, with either a nucleic acid molecule according to the invention and determining hybridization under stringent conditions of said nucleic acid molecule to a target nucleic acid molecule for determination of presence of a nucleic acid molecule, or using antibodies, preferably monoclonal antibodies raised against the polypeptide according to the present invention. Also a quantitative detection of the gene may be performed by PCR techniques, preferably by the use of quantitative RT-PCR using, e.g., the LightCycler™ of Roche Diagnostics GmbH, DE.

[0029] Since the Serpin gene seems to be associated with probiotic activity of bacterial strains, the present nucleic acid and/or polynucleotide may likewise be utilized for searching for additional strains exhibiting probiotic activities.

[0030] To determine, whether a given bacterial strain may be apt, the approximate amount of hybridization of the nucleic acid with the target nucleic acid or nucleic acids of the bacterial strain is determined. The approximate amount of hybridization may easily be determined qualitatively by e.g. visual inspection upon detecting hybridization. For example, if a gel is used to resolve labelled nucleic acid which hybridizes to target nucleic acid in the sample, the resulting band can be inspected visually. Likewise, as with the use of antibodies FACS may be utilized for a quantitative measurement.

[0031] The nucleic acid or a polypeptide according to the present invention may be utilized for the identification, characterization and/or purification of molecules affecting biological process associated with the activity of serine proteases. Exemplary biological processes, in which serine proteases are involved comprise blood coagulation, fibrinolysis, immune reactions, complement activation, inflammatory responses extracellular matrix turnover, cell migration and prohormone activation, cancer metastasis.

[0032] Likewise, the nucleic acid or polypeptide according to the present invention may also be used for the development of molecules eventually suitable in the treatment and/or diagnosis of disease states, which involve the activity of serine proteases. Once the interaction of the present Serpin with the target proteases has been understood, agonists or antagonists to the proteases, but likewise agonists or antagonists to Serpins may be devised. As mentioned above, non-limiting examples for disease states considered by the present invention are improper blood coagulation, improper fibrinolysis, immune reactions, complement activation, inflammatory responses extracellular matrix turnover, cell migration and prohormone activation, cancer metastasis.

[0033] The use of various model systems or structural studies will enable the development of specific agonists and antagonists useful in regulating the function of the present Serpin and the proteases to which it binds. It may be envisaged that these can be peptides, mutated ligands, antibodies or other molecules able to interact with the present Serpin.

[0034] The nucleic acid according to the present invention may in general be utilized for the synthesis of the polypeptide for large scale production thereof, by expressing a nucleic acid according to the present invention or a vector containing such a nucleic acid in a suitable host under conditions suitable for the expression of the polypeptide and collecting and purifying the polypeptide.

[0035] The following examples illustrate the invention without limiting it thereto.

Example 1

40 Isolation of the Serpin gene

[0036] In the line of sequencing the genome of the *Bifidobacterium longum* strain BL29 by the method of directed sequencing after fluorescent automated sequencing of the inserts of clones and assembling of these sequences of nucleotide fragments (inserts) by means of software programmes, an open reading frame has been found. To achieve that, fragments of the genome were created, ligated into suitable vectors for amplification and propagation and the corresponding fragments have been sequenced. Overlaps and the final arrangement of the fragments, the nucleotide sequence thereof, were assessed by the aid of appropriate softwares.

[0037] Protein and/or nucleic acid sequence homologies have been evaluated using the following algorithms:

- 50 (1) BLASTP: Compares an amino acid query sequence against a protein sequence database
- (2) BLASTN: Compares a nucleotide query sequence against a nucleotide sequence database
- (3) BLASTX: Compares a nucleotide query sequence translated in all reading frames against a protein sequence database
- (4) TBLASTN: Compares a protein query sequence against a nucleotide sequence database dynamically translated in all reading frames

A modest overall homology to the known murine Serpin α -2-antiplasmin of about 43 % has been noted. Yet the present polypeptide shows a homology of about 63 % with the reactive site loop (RSL) thereof.

Example 2

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Cloning of the Bifidobacterial Serpin gene and isolation of the polypeptide

[0038] The nucleic acid encoding the putative Bifidobacterial Serpin deleted from its signal peptide, was cloned into the E. coli expression vector pDEST 17 (Invitrogen Life Technologies) and the corresponding protein was produced

10 as a 6-His tagged fusion protein in E. coli (Janknecht R et al. (1991) : Proc. Natl. Acad. Sci. USA:88(20) pp 8972-8976) according to common techniques.

[0039] The 6-His tagged protein was purified to homogeneity by metal affinity chromatography on a nickel-nitrilotriacetic acid matrix (Ni-NTA from Quiagen) according to the instructions described in the QIA-Expressionist Handbook from Quiagen, and used for production of functional studies as well as for production of polyclonal antibodies in rabbits.

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Example 3

Potential anti-inflammatory activity of a serpin-like protein, expressed in BL29

20 [0040] The polypeptide as isolated in example 2 has been used in a haemolytic assay *in vitro* in order to determine its activity as a Serpin.

[0041] To this end, 50 μ l of successive doubling dilution of human AB serum (Sigma) were mixed in a 96 microliter well plate with 50 μ l of a 1.7 % sheep antibody-activated sheep erythrocyte suspension. To assess anti-haemolytic activity of the recombinant Serpin, 1 μ g of the recombinant protein was added to each well and the mixture was incubated for 1 hr at 37°C. The plate was then centrifuged to pellet intact cells and cell debris and the supernatant was transferred to a new plate. Haemolytic activity was estimated by measuring haemoglobin release as absorbance at 450 nm.

25 [0042] Bifidobacterium Serpin activity was compared to human serpin α 1-antitrypsin (5 μ g/well). Distilled water was used as positive control to obtain 100% erythrocyte lysis and human inactivated AB serum as negative control (no lysis due to complement inactivation).

30 [0043] The results clearly indicate that recombinant Serpin from Bifidobacteria inhibited human red blood cell lysis to a similar extent as human α 1-antitrypsin, a known serine-protease inhibitor.

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SEQ. Id. No. 1

5 Serpin nucleotide

10 ATGAGCGAGCAACTGATGGAACAGTACCGGTTGCGCGGACAACGCAAATGCCGTACGCT
 15 TGTATCGCCGCCATCGTACAGTAGTGCTTGCCTGCCGTCGCCGGCGCGTATGGTGG
 20 ACGGCCGGCGATGGCAGCGCATTGGTCGCAATATGTTCAAGCCGAAGGCCACGCCCTGCC
 25 ACGCAGCCGGTAGTCAACAGCACCGAACCTCGCCTACCGCACCGCACCGGAATTCTG
 30 GCGATGGAAGCCGGCGACCGAGGGCACCGGAATGTGAACACTACTCTCCTGCTTCGATGTGG
 35 ATGGCGTTGGCCATGCCCGCGCAGGGGCCAATGGCACCGACCCGCTCGCAACTGAACGAA
 40 CTGCTGGCTCCGGTTGCTGACCGATAGCGACTACCAATCGCTGCTAAGTTGATCAAC
 45 GGGCAATATTGGGGCGAAATCCGAGATGAGCGCCGCAACTCGCTGTGGATTGATGAC
 50 GACTACTCTCTGCCCAGCGATTACCAATCCACCGTCAAGAAGATGTTGAGGCCGAAGTC
 55 ACCACGTTACCGTTGACCGATCAGGCCGCCAAGATGTCGATTGGATTGCCAAGCAT
 60 ACGAATGGTCGCTCAAGCCGAAGATCACGCTGCGTGACCGTGAAGTCCTGTCCATCATC
 65 AACACCGTCTATCGGGATGGCGCTGGAGGATCCGTTGAGAGCAGTCCACCGGCAAC
 70 GGCACCTCCACGGCGAAGCCGGAGATGCTCAGGTGCCGATGATGCACCAAGACCTTCAGC
 75 CAAATGGCTTACGGACATGATGAGTACAACACTGGCAGCGGGTGGAGATTCCGTTGAC
 80 AACGGCGCAATCTGGCCATCGTGTGCCGCCAAGGGCATTGACGAGTTGGCCGGC
 85 GATGCCGAGAAGCTCAGTGGCGTTGGTACATGCTGACGGCATCCCTGGCGAGGGC
 90 GCAATGGTTGCCTCGCGACAGTATGCCGGCTGGCGTCTCCGTCACACTCGGTATG
 95 GTGAACGTACCGTACCGCGATTACCATCGACAGCATGTTGACTCGGAAGGCCACCATC
 100 AAGGCATTGCAAAACTGGGGGTGACCGATGCGTTGAGTCAGGCGACGCCACTTCACC
 105 AAGATGATCGACACCGGTTGACGGCGAGAACCTGTATATCGGCTGATTGCAAGGC
 110 ACGCGCATCGAGGTGAACGAAGCCGGCCAAGGCCATGTCCTCACCAAGGTGGCGCA
 115 GACTCCGTTAGCGCGCCGGTGGACAACGTCGAGTTACGGTGGATGCCATTCTGTAT
 120 TCGTACGTACCCCGGACGGCATAACCAATTATCATCGGTGCGGTGCGAACCTCGGCGGA
 125 GTCGGTGGAGAAAAC

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50

55

SEQ. ID. No. 2

5 Serpin protein Sequence

MetSerGluGlnLeuMetGluGlnTyrArgLeuArgGlyGlnArgLysCysArgAsnAla
 10 CysIleAlaAlalleValThrValValLeuValLeuAlaValAlaGlyGlyValTrpTrp
 ThrAlaGlyAspGlySerAlaLeuValArgAsnMetPheLysProLysAlaThrProAla
 15 ThrGlnProValValAsnSerThrAlaThrPheAlaTyrArgThrAlaProGluPheLeu
 AlaMetGluAlaGlyAspArgGlyThrGlyAsnValAsnTyrSerProAlaSerMetTrp
 MetAlaLeuAlalleAlaAlaGlnGlyAlaAsnGlyThrThrArgSerGlnLeuAsnGlu
 20 LeuLeuGlySerGlySerLeuThrAspSerAspTyrGlnSerLeuLeuSerSerIleAsn
 GlyGlnTyrSerGlyAlaLysSerGluMetSerAlaAlaAsnSerLeuTrpIleAspAsp
 AspTyrSerLeuAlaSerAspTyrGlnSerThrValLysMetPheGluAlaGluVal
 25 ThrThrLeuProPheAspAspGlnAlaAlaAlaLysMetSerAspTrpIleAlaLysHis
 ThrAsnGlySerLeuLysProLysIleThrLeuArgAspArgGluValLeuSerIleIle
 30 AsnThrValTyrAlaAspGlyArgTrpLysAspProPheGluGluGlnSerThrGlyAsn
 GlyThrPheHisGlyGluAlaGlyAspAlaGlnValProMetMetHisGlnThrPheSer
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 35 AspAlaGluLysLeuSerTrpAlaPheGlyThrCysSerThrAlaSerLeuGlyGluGly
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 LysAlaPheGluLysLeuGlyValThrAspAlaPheSerAlaGlyAspAlaAspPheThr
 LysMetIleAspThrGlySerHisGlyGluAsnLeuTyrIleGlySerIleLeuGlnGly
 45 ThrArgIleGluValAsnGluAlaGlyAlaLysAlaMetSerPheThrLysValGlyAla
 AspSerValSerAlaProValAspAsnValGluPheThrValAspArgProPheLeuTyr
 50 SerTyrValThrProAspGlyIleProLeuPhelleGlyAlaValArgAsnLeuGlyGly
 ValGlyGlyGluAsn

5 acgcgcacgc aggtgaacga agccggcgcc aagccatgt cttcaccaa gtcggcgca 1260
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 gtcggtgag aaaaac 1395

10 <210> 2
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 <213> **Bifidobacterium longum**

15 <400> 2

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1					5					10				15	

20 Cys Arg Asn Ala Cys Ile Ala Ala Ile Val Thr Val Val Leu Val Leu
 20 25 30

Ala	Val	Ala	Gly	Gly	Val	Trp	Trp	Thr	Ala	Gly	Asp	Gly	Ser	Ala	Leu
35								40					45		

25 Val Arg Asn Met Phe Lys Pro Lys Ala Thr Pro Ala Thr Gln Pro Val
 50 55 60

Val	Asn	Ser	Thr	Ala	Thr	Phe	Ala	Tyr	Arg	Thr	Ala	Pro	Glu	Phe	Leu
65				70					75				80		

30 Ala Met Glu Ala Gly Asp Arg Gly Thr Gly Asn Val Asn Tyr Ser Pro
 85 90 95

Ala	Met	Glu	Ala	Gly	Asp	Arg	Gly	Thr	Gly	Asn	Val	Asn	Tyr	Ser	Pro
35								40					45		

35 Ala Ser Met Trp Met Ala Leu Ala Ile Ala Ala Gln Gly Ala Asn Gly
 100 105 110

Ala	Ser	Met	Trp	Met	Ala	Leu	Ala	Ile	Ala	Ala	Gln	Gly	Ala	Asn	Gly
40								45					50		

40 Thr Thr Arg Ser Gln Leu Asn Glu Leu Leu Gly Ser Gly Ser Leu Thr
 115 120 125

Asp	Ser	Asp	Tyr	Gln	Ser	Leu	Leu	Ser	Ser	Ile	Asn	Gly	Gln	Tyr	Ser
45						130			135			140			

45 Gly Ala Lys Ser Glu Met Ser Ala Ala Asn Ser Leu Trp Ile Asp Asp
 145 150 155 160

Gly	Ala	Lys	Ser	Glu	Met	Ser	Ala	Ala	Asn	Ser	Leu	Trp	Ile	Asp	Asp
50															

50 Asp Tyr Ser Leu Ala Ser Asp Tyr Gln Ser Thr Val Lys Lys Met Phe
 165 170 175

Glu	Ala	Glu	Val	Thr	Thr	Leu	Pro	Phe	Asp	Asp	Gln	Ala	Ala	Lys
55								180			185		190	

Met Ser Asp Trp Ile Ala Lys His Thr Asn Gly Ser Leu Lys Pro Lys
 195 200 205

5 Ile, Thr Leu Arg Asp Arg Glu Val Leu Ser Ile Ile Asn Thr Val Tyr
 210 215 220

10 Ala Asp Gly Arg Trp Lys Asp Pro Phe Glu Glu Gln Ser Thr Gly Asn
 225 230 235 240

15 Gly Thr Phe His Gly Glu Ala Gly Asp Ala Gln Val Pro Met Met His
 245 250 255

20 Gln Thr Phe Ser Gln Met Ala Tyr Gly His Asp Glu Tyr Asn Thr Trp
 260 265 270

25 Gln Arg Val Glu Ile Pro Phe Asp Asn Gly Gly Asn Leu Ala Ile Val
 275 280 285

30 Leu Pro Ala Glu Gly His Phe Asp Glu Leu Ala Gly Asp Ala Glu Lys
 290 295 300

35 Leu Ser Trp Ala Phe Gly Thr Cys Ser Thr Ala Ser Leu Gly Glu Gly
 305 310 315 320

40 Ala Met Gly Cys Ala Ala Asp Ser Met Pro Gly Trp Gly Val Ser Val
 325 330 335

45 Asn Ser Val Met Val Asn Val Thr Leu Pro Arg Phe Thr Ile Asp Ser
 340 345 350

50 Met Phe Asp Ser Glu Ala Thr Ile Lys Ala Phe Glu Lys Leu Gly Val
 355 360 365

55 Thr Asp Ala Phe Ser Ala Gly Asp Ala Asp Phe Thr Lys Met Ile Asp
 370 375 380

60 Thr Gly Ser His Gly Glu Asn Leu Tyr Ile Gly Ser Ile Leu Gln Gly
 385 390 395 400

65 Thr Arg Ile Glu Val Asn Glu Ala Gly Ala Lys Ala Met Ser Phe Thr
 405 410 415

70 Lys Val Gly Ala Asp Ser Val Ser Ala Pro Val Asp Asn Val Glu Phe
 420 425 430

75 Thr Val Asp Arg Pro Phe Leu Tyr Ser Tyr Val Thr Pro Asp Gly Ile

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440

445

5 Pro Leu Phe Ile Gly Ala Val Arg Asn Leu Gly Gly Val Gly Gly Glu
450 455 460

10 Asn
465

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Claims

1. A nucleic acid as identified by SEQ ID. NO. 1 or parts or variants thereof coding for a functional polypeptide, which variants have a homology to the SEQ ID. No. 1 of about 75 %, preferably 80 %, more preferably 85 %, even more preferably 90 % even more preferred 95 %.

5 2. A polypeptide as identified by SEQ ID. NO. 2 or functional parts or variants thereof, that have a degree of homology to the said SEQ. ID. No. 2 of about 75 %, preferably 80 %, more preferably 85 %, even more preferably 90 % even more preferred 95 %.

10 3. A vector containing a nucleic acid according to claim 1.

4. A host cell containing one or more copies of a nucleic acid according to claim 1 or a vector according to claim 3.

15 5. A host cell according to claim 4, wherein a polypeptide according to claim 2 is expressed.

6. Use of a nucleic acid or a polypeptide according to any of the claims 1 or 2 for the identification, characterization and/or purification of molecules affecting a biological process associated with the activity of serine proteases.

20 7. Use of a nucleic acid or a polypeptide according to any of the claims 1 or 2 for the development of molecules for the treatment and diagnosis of disease states, involving the activity of serine proteases.

8. Use of a nucleic acid according to claim 1 or a polypeptide according to claim 2, for the preparation of a carrier for the treatment and/or prevention of disease states involving the activity of serine proteases.

25 9. The use according to claim 8, wherein the carrier is a medicament, a food product or a food supplement.

10. The use according to any of the claims 6 to 9, wherein the biological process or the disease states are selected from the group consisting of blood coagulation, fibrinolysis, immune reactions, complement activation, inflammatory responses extracellular matrix turnover, cell migration and prohormone activation, cancer metastasis.

30 11. A method of producing a polypeptide according to claim 2, which comprises expressing a nucleic acid according to claim 1 or a vector according to claim 3 in a suitable host and purifying the polypeptide obtained.

35 12. A method for detecting probiotic strains, which comprises screening micro-organisms with a nucleic acid according to claim 1 or an antibody directed to a polypeptide according to claim 2, for the presence of the said nucleic acid or the said polypeptide and determining the presence of a Serpin in the micro-organisms.

40 13. A method for producing a micro-organism expressing or over-expressing the Serpin encoded by a nucleic acid according to claim 1, which comprises, transforming a micro-organism with a nucleic acid according to claim 1 and expressing the gene encoding the Serpin polypeptide.

45 14. A food or pharmaceutical product, containing a nucleic acid according to claim 1, a polypeptide according to claim 2, a vector according to claim 3 or a host cell according to any of the claims 4 or 5.

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